

Learning Multiple Evolutionary Pathways from Cross-sectional Data

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ABSTRACT

We introduce a mixture model of trees to describe evolutionary processes that are characterized by the accumulation of permanent genetic changes. The basic building block of the model is a directed weighted tree that generates a probability distribution on the set of all patterns of genetic events. We present an EM-like algorithm for learning a mixture model of K trees and show how to determine K with a maximum likelihood approach. As a case study we consider the accumulation of mutations in the HIV-1 reverse transcriptase that are associated with drug resistance. The fitted model is statistically validated as a density estimator and the stability of the model topology is analyzed. We obtain a generative probabilistic model for the development of drug resistance in HIV that agrees with biological knowledge. Further applications and extensions of the model are discussed.

Categories and Subject Descriptors

J.3 [Life and Medical Sciences]: Biology and genetics;
I.5.1 [Pattern Recognition]: Models—*Statistical*

General Terms

Algorithms

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Keywords

Mixture models, Tree models, Bayesian networks, EM algorithm, HIV drug resistance, Mutational pathways

1. INTRODUCTION

Despite the introduction of 18 different drugs that inhibit replication of human immunodeficiency virus type 1 (HIV-1), therapeutic success is still limited. A major factor contributing to therapy failure even of modern combination therapies (highly active antiretroviral therapy, HAART) is the virus' ability to escape from drug pressure by developing drug resistance [28, 35]. This escape mechanism is based on HIV's high rates of replication and mutation. Residual viral reproduction under therapy allows for generating genetic variants that have a selective advantage under drug pressure. Eventually, some of these mutants may replicate as well or nearly as well as a wild type virus and thus lead to viral rebound.

Considerable work has been carried out on characterizing the relationship between genetic changes in the viral drug targets and phenotypic drug resistance. Many single mutations have been linked to resistance against one or more drugs [32]. Mutational patterns conferring resistance have been identified by several statistical and machine learning methods [2, 4, 31]. Some computational approaches make use of this (either data-derived or expert) knowledge in order to find optimal drug combinations based on sequence information on the viral drug targets [1, 25]. In particular, it has been shown that predictions of clinical response can benefit from exploring possible mutational pathways of the virus.

However, much less is known about *how* resistance-associated mutations accumulate. Some mutations are known to occur preferentially in clusters [3, 15, 37], but the order of accumulation is usually unknown. Only a few studies based on longitudinal (time series) data have revealed directed dependencies between mutations [5, 27]. This type of analysis is not practical for many different drugs or even drug combinations, because large longitudinal samples from patients under the same therapeutic regimen are difficult to obtain. As an alternative, we propose a method for estimating mutational pathways from cross-sectional data (i.e. data from different patients at different time points), which are much more abundant.

We develop the technique in a general setting and consider the development of HIV-1 drug resistance to the nucleoside reverse transcriptase inhibitor zidovudine as a test case.

1.1 Zidovudine Resistance

Zidovudine (AZT) has been approved for clinical use in 1987 as the first anti-HIV drug. Soon after approval mutations in HIV-1 reverse transcriptase (RT) have been found that decrease susceptibility to the drug and develop within a few month of therapy [23, 24]. The most common RT mutations (“classical zidovudine mutations”) that develop under zidovudine therapy are M41L¹, D67N, K70R, L210W, T215F/Y, and K219E/Q. Other mutations such as the multi-nucleoside resistance mutations V75I, F77L, F116Y, Q151M and the two amino acids insertion after position 69 are less frequent and typically occur under prolonged combination therapies containing two or more nucleoside RT inhibitors. K70R and T215F/Y are generally the first mutations to occur causing 4- to 8- and 10- to 16-fold zidovudine resistance *in vitro*, respectively [5, 22]. The double mutant 41L+215F/Y causes already 60- to 70- fold resistance. 41L, 215F/Y and 210W tend to occur together (215-41 pathway) as well as 70R and 219E/Q (70-219 pathway). Substitution M41L may also appear first, albeit less frequently. However, in contrast to the 70R+215F/Y double mutant, the 41L+70R co-occurrence is hardly ever observed. This discrepancy is explained by a replication defect of the 41L+70R intermediate [19].

In general, the evolution of drug resistance is driven by several factors including

- nucleotide substitution patterns such as transition/transversion bias [21],
- viral fitness (replication capacity of the virus),
- drug resistance (decreased drug susceptibility).

Under therapy the viral population is exposed to a strong selective pressure. Mutations almost always arise one at a time and each single advantageous mutation must be fixed into the population. In consequence, relatively few evolutionary pathways lead from the wild type to a highly resistant, well replicating mutant as compared to the large number of possible mutational patterns [16].

1.2 Outline

We describe the evolution of drug resistance as the accumulation of permanent genetic changes. Our model of the stochastic evolutionary process aims at identifying directed dependencies between mutational events. The basic building block of the model is a directed tree. Vertices of the tree represent binary random variables, each indicating the occurrence of an event (mutation). Edge weights represent conditional probabilities between events with the constraint that a child event does not occur whenever the parent event has not occurred. These restricted Bayesian tree models have been pioneered by Desper *et al.* in the context of oncogenesis [9]. In section 2 we recall basic model properties and an efficient combinatorial algorithm for tree reconstruction from observed (cross-sectional) patterns of events.

The tree models provide a detailed and interpretable description of the process of accumulating genetic changes. They represent a considerable improvement over independence or linear path models. Nevertheless, we will see that

¹We use the syntax $a x b$ (or simply $x b$) to denote amino acid substitutions in RT, where a is the amino acid in the reference strain HXB2 at position x and b the mutated residue.

the special tree structure fits only certain subgroups of the data. We interpret this shortcoming as indicating that the data has been generated by more than one (tree-like) process. Therefore, we introduce the broader class of mixture models of trees. Ideally, we would like to identify multiple evolutionary processes acting on the same gene (or genome), each process in one specialized component of the mixture model. In particular, we will introduce a “noise component” that includes all otherwise unexplained samples. We define these mixture models in section 3 and present an EM-like algorithm for learning structure and parameters of the model from data. In section 4 we illustrate how model selection (choosing the optimal number of trees) can be performed in a maximum likelihood fashion. We compare the mixture model for the development of zidovudine resistance with biological knowledge.

In section 5 we present cross-validation and bootstrap methods for the validation of our model. Section 6 discusses further applications of the method, related work and open questions.

2. MUTAGENETIC TREES

2.1 Data Representation

We consider ℓ different events $\{1, \dots, \ell\}$, including a special “null event” that has initially occurred in all samples. A pattern x_i of events is represented by a row vector of indicator variables of length ℓ :

$$x_i = (x_{i1}, \dots, x_{i\ell}),$$

$$x_{ij} = \begin{cases} 1, & \text{if event } j \text{ has occurred in sample } i \\ 0, & \text{else.} \end{cases}$$

Thus, a set of N observed patterns is represented by the binary matrix

$$X = (x_{ij})_{\substack{1 \leq i \leq N \\ 1 \leq j \leq \ell}}.$$

We denote by $\Omega = 2^{\{1, \dots, \ell\}}$ the set of all possible patterns of length ℓ .

For the case of HIV-1 zidovudine resistance we consider a set of $N = 364$ samples derived from previously untreated patients under zidovudine mono-therapy as available from the Stanford HIV Drug Resistance Database [30]. No resistance-associated mutations other than the six classical zidovudine mutations are present in this data set. Thus, the set of $\ell = 7$ events comprises 41L, 67N, 70R, 210W, 215F/Y, and 219E/Q, plus the initial state characterized by M41, D67, K70, L210, T215, and K219 and referred to as the “wild type”. The data set contains 35 different mutational patterns. The null pattern representing the wild type is observed in 115 samples.

We reconstruct dependencies between events from the joint probabilities between all pairs of events, which can be estimated reliably from moderately large data sets.

2.2 Definition

To describe the ordered accumulation of genetic changes, we consider directed trees over the set of events, where each edge is weighted with the conditional probability of the child event given that the parent event has occurred. Formally, a

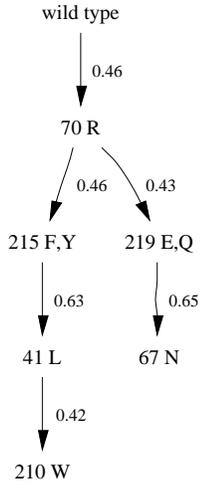


Figure 1: Mutagenetic tree for the development of zidovudine resistance. Nodes are labeled with resistance-associated mutations in the HIV-1 reverse transcriptase, edge labels represent conditional probabilities between mutational events.

*mutagenetic tree*² $\mathcal{T} = (V, E, r, p)$ consists of a set of vertices V representing events, a set of edges E , a special vertex $r \in V$, and a map $p : E \rightarrow [0, 1]$ such that:

- (V, E) is a branching, i.e. a digraph whose underlying undirected graph is a forest, and each vertex has at most one entering edge.
- The vertex r represents the null event and has no entering edge.
- For all edges $e = (j_1, j_2) \in E$,
 - $p(e) = \Pr(j_2|j_1)$ is the conditional probability of event j_2 given that event j_1 has occurred,
 - $p(e) > 0$ (if $p(e) = 0$, we can delete e from E),
 - $p(e) < 1$ if e leaves the root (if $\Pr(j_2|r) = 1$, events j_2 and r can be merged).

Note that a mutagenetic tree can have more than one connected component. However, most of the time we will be concerned only with the arborescence (connected branching) containing the special root vertex r .

Figure 1 shows a mutagenetic tree for the development of resistance to zidovudine.

A mutagenetic tree induces a probability distribution on the set Ω of all possible mutational patterns as follows. Draw each edge independently from E with probability $p(e)$. Then the set of vertices that are reachable from the root is the outcome of the experiment.

2.3 Tree Reconstruction

Desper *et al.* have shown how to reconstruct the mutagenetic tree from all pairwise joint probabilities of events [9]. Consider the complete digraph $G = (V, V \times V, w)$ on the set of vertices V corresponding to the events with weights

$$w(j_1, j_2) = \log \Pr(j_1, j_2) - \log(\Pr(j_1) + \Pr(j_2)) - \log \Pr(j_2),$$

²We follow the notation of Desper *et al.* who call these tree models in the context of oncogenesis *oncogenetic trees* [9].

where $\Pr(j)$ denotes the marginal probability of event j and $\Pr(j_1, j_2)$ the joint probability of events j_1 and j_2 . Then the mutagenetic tree is the branching in G that maximizes the sum of its edge weights. The maximum weight branching can be computed in $O(|V||E|)$ time by Edmonds’ branching algorithm [7, 11, 20, 34].

In practice, we do not know the joint probabilities of events, but have to estimate them from the data. For sufficiently many samples the above algorithm will reconstruct the correct mutagenetic tree with high probability (see [9] for proofs and a quantitative version of this statement).

Finally, if the observed sample does not come from a distribution generated by a mutagenetic tree, we hope that the reconstructed tree captures many of the strong dependencies (causality flows) between events.

2.4 Likelihood Computation

Given a mutagenetic tree $\mathcal{T} = (V, E, r, p)$, the likelihood of a pattern x of events is the probability that \mathcal{T} generates x : $L(x|\mathcal{T}) = \Pr(x|\mathcal{T})$. Let $S \subseteq V$ be the set of events specified by x . If there is a subset $E' \subseteq E$ such that S is the set of all vertices reachable from r in the subtree (V, E') , then x can be generated by \mathcal{T} , and the likelihood is given by

$$L(x|\mathcal{T}) = \prod_{e \in E'} p(e) \cdot \prod_{e \in (S \times V \setminus S)} (1 - p(e)).$$

If there is no such edge subset, the topology of \mathcal{T} does not allow for generating x and hence $L(x|\mathcal{T}) = 0$. For example, for the mutagenetic tree displayed in Figure 1 and the pattern defined by the mutations 70R and 219Q we find

$$L(x|\mathcal{T}) = 0.46 \cdot 0.43 \cdot (1 - 0.46) \cdot (1 - 0.65) = 0.037,$$

whereas the pattern composed of 215Y and 41L can not be generated by that tree.

The likelihood computation can be done efficiently by traversing the mutagenetic tree in a breadth first search starting from r . Note that connected components of \mathcal{T} other than the arborescence rooted at r do not affect the likelihood of a pattern.

We call \mathcal{T} a star, if all edges $e \in E$ leave the root vertex r . The star topology models events as being independent of each other. In terms of the likelihood a star is characterized as follows.

LEMMA 1. *A mutagenetic tree is a star, if and only if all 2^{ℓ} possible patterns of events have positive likelihood.*

PROOF. If \mathcal{T} is a star,

$$L(x|\mathcal{T}) = \prod_{\{j|x_j=1\}} \Pr(j) \cdot \prod_{\{j|x_j=0\}} (1 - \Pr(j)) > 0,$$

since $\Pr(j) = \Pr(j|r) \in (0, 1)$ by definition.

If \mathcal{T} is not a star, there is at least one edge (j_1, j_2) with $j_1 \neq r$ and any pattern with $x_{j_1} = 0$ and $x_{j_2} = 1$ has likelihood zero. \square

Thus, for noisy real world data the assumption of a tree topology will generally be too strict in the likelihood sense. Moreover, the estimated mutagenetic tree for the development of zidovudine resistance (Figure 1) does not capture all of the known pathways. Indeed, the tree topology implies that M41L and T215F/Y can only occur *after* K70R despite the fact that the 215-41 pathway is also observed

in the absence of K70R. Consequently, one third of the observed mutational patterns have likelihood zero under the estimated mutagenetic tree.

To overcome these limitations we consider the broader class of mixture models of mutagenetic trees.

3. MIXTURE MODELS

3.1 Definition

Suppose that Y_1, \dots, Y_K are multivariate discrete random variables with range Ω that are distributed according to mutagenetic trees

$$\mathcal{T}_k = (V, E_k, r, p_k), \quad k = 1, \dots, K,$$

respectively. Let $\Delta_1, \dots, \Delta_K \in \{0, 1\}$ be binary random variables with $\Pr(\Delta_k = 1) = \alpha_k$. We call the model

$$\mathcal{M} = \sum_{k=1}^K \alpha_k \mathcal{T}_k \quad \text{with} \quad \alpha_k \in [0, 1] \quad \text{and} \quad \sum_{k=1}^K \alpha_k = 1,$$

that generates the random variable $Y = \sum_{k=1}^K \Delta_k Y_k$, a *K-mutagenetic trees mixture model*.

Thus, the likelihood of a pattern of events x in the mixture model is

$$L(x|\mathcal{M}) = \sum_{k=1}^K \alpha_k L(x|\mathcal{T}_k).$$

Throughout we will consider mixture models that have a special structure in the first mutagenetic tree \mathcal{T}_1 . We assume that, in addition to different pathways of accumulation of events, there is a certain probability β of any event occurring spontaneously independent of all other events. Thus, \mathcal{T}_1 is a star with $p(e) = \beta$ for all $e \in E_1$. \mathcal{T}_1 can be regarded as the noise component of the model. Including a star in the mixture model ensures that all patterns of events have positive likelihood.

3.2 EM-like Learning Algorithm

Given the number of trees K , we want to reconstruct a K -mutagenetic trees mixture model from observed patterns X . This task would be easy, if we knew for each pattern of events from which component(s) of the model it has been generated: We would apply K times the reconstruction technique for a single tree. However, this information is missing and we have to estimate it from the data, too. This procedure results in an algorithm similar to an EM algorithm [8].

Our goal is to find mutagenetic trees $\mathcal{T}_1, \dots, \mathcal{T}_K$ and mixture parameters $\alpha_1, \dots, \alpha_K$ that maximize the log-likelihood of the data, which can be written as

$$\sum_{i=1}^N \log \sum_{k=1}^K \alpha_k L(x_i|\mathcal{T}_k),$$

if the x_i are independent. The *responsibility* of model component k for sample x_i is defined as

$$\gamma_{ik} = \Pr(\Delta_k = 1|\mathcal{M}, x_i).$$

Let $N_k = \sum_{i=1}^N \gamma_{ik}$ be the weighted number of samples generated by \mathcal{T}_k . In an iterative fashion, we estimate γ (E step) and \mathcal{M} (M step) from the data.

INPUT:

- Patterns of events $X = (x_{ij})_{\substack{1 \leq i \leq N \\ 1 \leq j \leq \ell}}$
- Number of mutagenetic trees $K \geq 2$

OUTPUT:

- K -mutagenetic trees mixture model $\sum_{k=1}^K \alpha_k \mathcal{T}_k$

PROCEDURE:

1. Guess initial responsibilities:

- (a) Run $(K - 1)$ -means clustering algorithm
- (b) Set responsibilities

$$\gamma_{ik} = \begin{cases} \frac{1}{2}, & \text{if } x_i \text{ is in cluster } k - 1 \\ \frac{1}{2(K-1)}, & \text{else.} \end{cases}$$

2. *M-like step*. Update model parameters:

Set $N_k = \sum_{i=1}^N \gamma_{ik}$ for all $k = 1, \dots, K$.

Let \mathcal{T}_1 be a star with edge weights

$$\beta = \frac{1}{\ell N_1} \sum_{j=1}^{\ell} \sum_{i=1}^N \gamma_{i1} x_{ij}.$$

For $k = 2, \dots, K$:

- (a) For all pairs of events (j_1, j_2) , $1 \leq j_1, j_2 \leq \ell$, estimate their joint probabilities

$$p_k(j_1, j_2) = \frac{1}{N_k} \sum_{i=1}^N \gamma_{ik} x_{ij_1} x_{ij_2}.$$

- (b) Compute the maximum weight branching T_k from the complete digraph with weights w derived from p_k .

- (c) Compute the mixture parameter $\alpha_k = \frac{N_k}{N}$.

3. *E step*. Compute responsibilities:

$$\gamma_{ik} = \frac{\alpha_k L(x_i|T_k)}{\sum_{m=1}^K \alpha_m L(x_i|T_m)}.$$

4. Iterate steps 2 and 3 until convergence.

Figure 2: EM-like algorithm for learning a K -mutagenetic trees mixture model from data.

Given an estimate of $\mathcal{M} = \sum_{k=1}^K \alpha_k \mathcal{T}_k$, we can estimate γ by

$$\gamma_{ik} = \frac{\alpha_k L(x_i|\mathcal{T}_k)}{\sum_{m=1}^K \alpha_m L(x_i|\mathcal{T}_m)}.$$

Given an estimate of γ , we update \mathcal{M} as follows. For the noise component ($k = 1$) we choose the star topology and estimate β as the rate of occurrence of any event in this component,

$$\beta = \frac{1}{\ell N_1} \sum_{j=1}^{\ell} \sum_{i=1}^N \gamma_{i1} x_{ij}.$$

For $k \geq 2$, we first estimate all joint probabilities between pairs of events within the k -th component:

$$p_k(j_1, j_2) = \frac{1}{N_k} \sum_{i=1}^N \gamma_{ik} x_{ij_1} x_{ij_2}.$$

Next, we reconstruct \mathcal{T}_k from p_k by solving the maximum weight branching problem as described in section 2.3. Edges with $p_k(j_1, j_2) < 0.01$ are previously deleted from the complete graph in order to avoid weakly connected components within one mutagenetic tree. Finally, the mixture parameters are updated by the equation

$$\alpha_k = \frac{N_k}{N} = \frac{1}{N} \sum_{i=1}^N \gamma_{ik}.$$

We iterate the E step and the M step until the log-likelihood function does not increase any more.

To run the algorithm we need initial values for the responsibilities γ_{ik} . The starting solution can be picked at random, but in general this strategy will yield poor results. The two common approaches to overcome this problem are either to sample many random starting solutions, or to identify a single promising initial solution. To limit computational costs we decided for the latter approach and perform an ordinary k -means clustering with $k = K - 1$ on the set of patterns using squared Euclidean distance as dissimilarity measure³ [17]. From the k -means cluster assignments we derive the initial responsibilities

$$\gamma_{ik} = \begin{cases} \frac{1}{2}, & \text{if sample } x_i \text{ belongs to cluster } k - 1 \\ \frac{1}{2(K-1)}, & \text{else.} \end{cases}$$

The procedure is summarized in Figure 2. It is differing from a true EM algorithm in the fact that the tree reconstruction step does not provide a maximum likelihood estimate. Thus, unlike with a true EM algorithm, our EM-like algorithm is not guaranteed to converge to a local maximum of the log-likelihood function. Nevertheless, we have not observed such deviating behavior on any real world data set so far.

4. MODEL SELECTION

There is still one model parameter that we do not know, namely the number of mutagenetic trees K in the mixture model. Our learning algorithm is efficient enough to perform model selection in a cross-validation setting. For the zidovudine resistance data, we have used 10-fold cross-validation in order to estimate the likelihood on unseen data for values of K between 1 and 10. Figure 3 shows the estimated likelihood as a function of K . We propose to apply the one-standard-error rule, i.e. to pick the most parsimonious model within one standard error of the maximum mean log-likelihood [17]. This yields $K = 3$, and the 3-mutagenetic trees mixture model for the development of zidovudine resistance in the HIV-1 RT is shown in Figure 4.

This model assigns 19% of the data to the noise compo-

³The starting solution for the k -means algorithm, i.e. the set of initial cluster centers, is a random subset of the data of size k . In all experiments, we have chosen the best k -means clustering (the one minimizing the within-cluster point scatter) obtained from 100 random starting solutions.

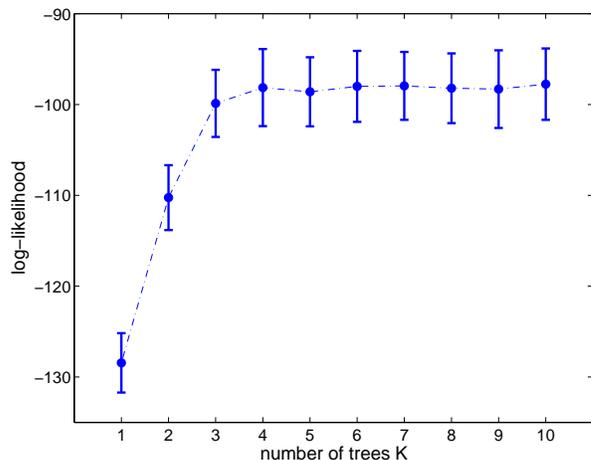


Figure 3: 10-fold cross-validation log-likelihood estimates for K -mutagenetic trees mixture models as a function of K .

nent⁴. These data are not necessarily free of any dependencies between events, but within the model class they are best explained by the independence assumption. 47% of the data are estimated to be generated by a linear path model that involves the 70-219 pathway followed by 67N and the 215-41 pathway. The remaining 34% of the data are assigned to a mutagenetic tree with initial event 215F/Y followed by either the 70-219 pathway or—with greater probability—the remainder of the 215-41 pathway, namely 41L and 210W.

In conclusion, the mixture model of mutagenetic trees captures all major established facts about the development of zidovudine resistance under zidovudine mono-therapy. Moreover, it provides a quantitative, generative probabilistic model of the accumulation of resistance-associated mutations.

5. VALIDATION

After comparing the estimated mixture model with biological knowledge, we now turn to quantitative approaches for model validation. We will derive measures of confidence regarding the mixture model as both a density estimator and a way of learning structural dependencies between events.

5.1 Goodness of Fit

Drawing a random sample from a K -mutagenetic trees mixture model $\mathcal{M} = \sum_{k=1}^K \alpha_k \mathcal{T}_k$ is straightforward. We first draw a uniform random number and decide according to the mixture parameters $\alpha = (\alpha_1, \dots, \alpha_K)$ which mutagenetic tree to use. In the selected tree we draw each edge $e \in E$ independently with probability $p(e)$. The sample consists of all events that are reachable from r in the induced subgraph.

We want to quantify how closely a trained mixture model reproduces the empirical probability distribution on $\Omega = 2^{\{1, \dots, \ell\}}$. To compare two histograms $H_1, H_2 \in \mathbb{N}^{2^\ell}$ we use the cosine distance, defined in terms of the angle spanned

⁴in the sense of the mixture model: in general, each sample is distributed over several components, cf. section 5.1.

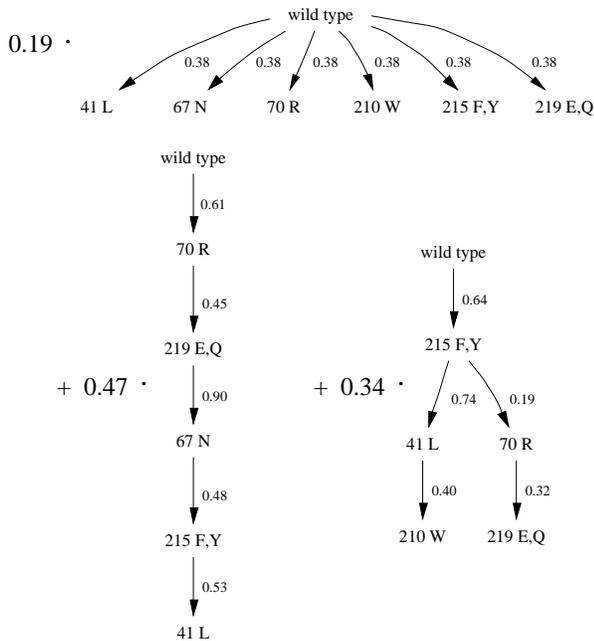


Figure 4: 3-Mutagenetic trees mixture model $\mathcal{M} = 0.19\mathcal{T}_1 + 0.47\mathcal{T}_2 + 0.34\mathcal{T}_3$ for the development of zidovudine resistance. Each tree is preceded by its weight α_k . The upper tree \mathcal{T}_1 is a star and represents the noise component. In the second tree we have omitted the connected component consisting of the single vertex with label 210W, and similarly 67N in the third tree.

by the histogram vectors as

$$\text{dist}(H_1, H_2) = 1 - \cos \angle(H_1, H_2) = 1 - \frac{\langle H_1, H_2 \rangle}{\|H_1\| \|H_2\|} \in [0, 1].$$

Using cross-validation we calculate for each partition of the data into training and test set the histogram distance between test data and

- 1) training data,
- 2) simulated data drawn from the optimal mixture model,
- 3) simulated data drawn from a single mutagenetic tree model,
- 4) simulated data drawn from a single star model with non-uniform edge weights.

The three models are estimated from the training data, and the size of the simulated sample equals that of the training data. The first histogram distance measures only the effect of finite sampling, whereas the other distances include losses that are due to imperfect model assumptions and/or parameter estimates. We compare the optimal mixture model with a single mutagenetic tree model and with a star model representing the null hypothesis of independence of events.

Figure 5 shows the distribution of all distances for 100 runs of 10-fold cross-validation each. Histograms generated from the estimated mixture model closely resemble the observed data. In contrast, both the single tree model and the independence assumption provide inferior model fits.

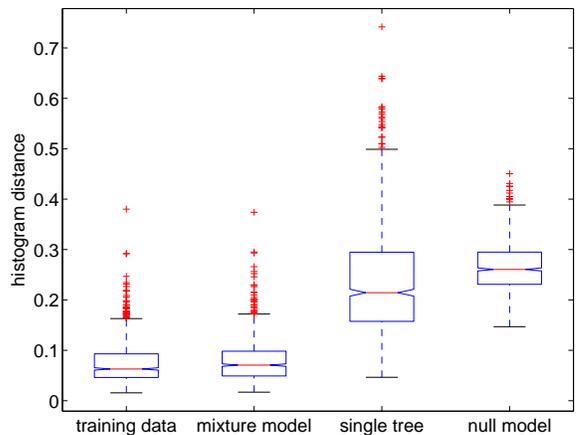


Figure 5: Box-plot of histogram distances from 100 10-fold cross-validation runs. From left to right: distances between histograms of test data and training data (first box), simulated data from the optimal mixture model (second box), simulated data from the single tree model (third box), simulated data from the null model (fourth box). Simulated data are drawn from the models that have been fitted on the training data. The null model refers to the independence assumption of events and is a single star with non-uniform edge weights.

likelihood			sample subset	
$L(x \mathcal{T}_1)$	$L(x \mathcal{T}_2)$	$L(x \mathcal{T}_3)$	fraction	description
> 0	> 0	> 0	31.6%	null patterns
> 0	> 0	$= 0$	30.2%	70-219 pathway
> 0	$= 0$	> 0	25.0%	215-41 pathway
> 0	$= 0$	$= 0$	13.2%	“noise”

Table 1: Distribution of samples among the components of the mixture model shown in Figure 4. The fraction of samples refers to the full data set of 364 samples.

Similar results are obtained with the L_2 and L_1 distance measures between histograms instead of the cosine distance.

In the same cross-validation runs we have determined the percentage of samples that remain unexplained by the non-trivial components of the mixture model. The mean percentage of samples with likelihood zero in all but the noise component was 13%. Thus, the mixture model maps 87% of the observed patterns onto the other identified mutagenetic trees. For the optimal model on the full data (Figure 4) it happens to be the case that the only pattern that can be generated by both non-trivial trees is the null pattern. We report in Table 1 the distribution of samples among the trees in detail.

5.2 Tree Stability

We have already interpreted the topology of the mutagenetic trees in detail. Here we use the bootstrap [12] in order to obtain an estimate of the dependence of the topology on sampling effects. Since there is no standard way of comparing two mixture models, we confine ourselves to analyzing the stability of each single mutagenetic tree. For a mixture model, we fix the responsibilities γ obtained from the EM-

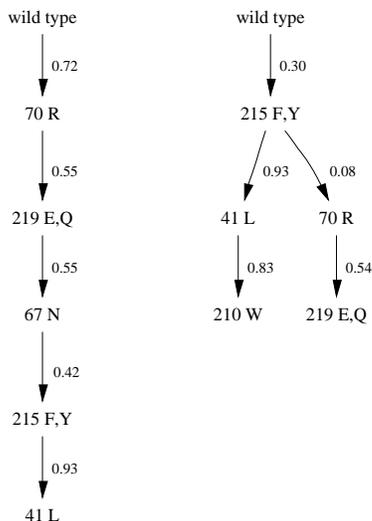


Figure 6: Bootstrap analysis of tree stability. Edge weights represent relative counts in 10,000 bootstrap samples. The two mutagenetic trees are the two non-trivial components of the mixture model in Figure 4.

algorithm. For tree \mathcal{T}_k , we resample with replacement each pattern of events x_i with probability γ_{ik} . From the bootstrap sample of size N a mutagenetic tree is reconstructed. As a test statistic we consider the relative count of each edge $e \in E_k$ in the bootstrap trees.

In Figure 6 we report for the zidovudine data these estimates based on 10,000 bootstrap samples. The two mutagenetic trees are the two non-trivial components of the mixture model displayed in Figure 4. We find strong support for most edges, particularly for 70R as initial event and for the dependencies 215F/Y \rightarrow 41L \rightarrow 210W and 70R \rightarrow 219E/Q \rightarrow 67N. The latter edge suggests that mutation D67N may be best explained as a late event in the 70-219 pathway.

6. DISCUSSION

We have presented mixture models of mutagenetic trees for modeling evolutionary processes that can be described as an accumulation of permanent genetic changes along multiple pathways. Application to the development of zidovudine resistance in the HIV-1 RT has shown that, compared to single mutagenetic trees, the class of mixture models provides both better density estimation of observed patterns of events and biologically more plausible models.

6.1 Applications

Further applications of the model include the accumulation of mutations associated with resistance to other antiretroviral drugs in protease and RT, the current drug targets of HAART. Together, these models may be helpful in designing treatment protocols that avoid the accumulation of cross-resistance conferring mutations so that clinicians do not run out of therapeutic options too early. However, since mono-therapies are obsolete, mutational pathways under combination therapy must be identified. It remains to be investigated if combination therapy pathways are a function of the pathways for the single drugs forming the com-

ination. If such a relationship can not be found, we may face another data scarceness problem induced by the large number of possible drug combinations.

As density estimators, the mixture models can also be used for classification. A common classification problem in the context of HIV drug resistance is to separate susceptible from resistant strains. With mixture models \mathcal{M}_{sus} trained on the susceptible and \mathcal{M}_{res} trained on the resistant subset, we consider the likelihood ratio

$$\frac{L(x|\mathcal{M}_{sus})}{L(x|\mathcal{M}_{res})}$$

to decide whether a pattern x is more likely to originate from the susceptible or the resistant subpopulation. Analysis of the model \mathcal{M}_{res} may reveal pathways leading to resistance independent of the applied drug pressure, including cross-resistance pathways induced by other drugs.

6.2 Model-based Clustering

The classical EM algorithm for learning Gaussian mixture models can be regarded as a soft version of K -means clustering [17]. For each model component, the EM algorithm assigns a responsibility to each sample, whereas the K -means clustering algorithm assigns each sample to exactly one of the K clusters. Likewise, we can easily modify our K -mutagenetic trees mixture model learning algorithm (Figure 2) to obtain a clustering algorithm. It suffices to store cluster assignments instead of responsibilities in the matrix γ :

$$\gamma_{ik} = \begin{cases} 1, & \text{if sample } x_i \text{ is in cluster } k \\ 0, & \text{else,} \end{cases}$$

and change the E step to

$$\gamma_{ik} = \begin{cases} 1, & \text{if } k = \arg \max_{1 \leq m \leq K} L(x_i|\mathcal{T}_m) \\ 0, & \text{else.} \end{cases}$$

This model-based clustering is useful in situations where pathways are known or suspected to be mutually exclusive.

6.3 Full ML Estimation

We would arrive at a true EM algorithm if we estimated the mutagenetic trees \mathcal{T}_k (Figure 2, step 2(b)) in a maximum likelihood (ML) fashion. However, finding the ML topology of a mutagenetic tree appears difficult in absence of a construction rule and in view of the large number of possible trees. Thus, heuristic search methods need to be applied, such as those used in ML phylogeny estimation [13].

6.4 Related Work

Chow and Liu have used unrestricted Bayesian tree models to approximate multivariate discrete probability distributions [6]. They show that solving the maximum weight spanning tree problem in the complete graph with edges between features (events) weighted by their mutual information provides a ML tree estimate. Maximum weight branchings have been proposed in a similar setting [18]. Our mutagenetic trees mixture models are similar in spirit to the work of Meilă and Jordan [26], who apply an EM algorithm to generalize the Chow-Liu algorithm to mixtures of trees. Friedman *et al.* have extended the Chow-Liu procedure for classification tasks [14].

Graph models related to mutagenetic trees have been developed for oncogenesis, where chromosomal losses and

gains are considered as events. The distance matrix between events defined by

$$-2 \log \Pr(j_1, j_2) + \log \Pr(j_1) + \log \Pr(j_2)$$

has been used as input for distance-based phylogeny methods like neighbor-joining [10]. The resulting phylogenetic tree represents events as leaves of the tree and groups closely related events together. Internal vertices are considered “hidden events” and do not have a direct interpretation.

A similar approach uses a ML estimation procedure for tree fitting [36]. A closed formula for the ML parameters of a tree is derived, while searching for the ML topology is done heuristically.

Finally, generalizing from tree models, directed acyclic graph (DAG) models have been proposed [29, 33]. Here vertices represent subsets of the set of events and an edge $\{j_1\} \rightarrow \{j_1, j_2\}$ represents the probability that j_1 occurs first and j_2 occurs second. Edges for larger subsets are defined similarly. For limited subset size ML estimation of model parameters is feasible.

Our mixture models have the same expressive power as DAG models. We expect DAG models to be more appropriate for highly connected networks of events, whereas mixture models may be more adequate to identify a few parallel or confluent pathways.

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